stable in the mass spectrometer. Catalytically active dehydroquinase was mixed with 3-dehydroquinate immediately prior to injection into the mass spectrometer. By using a large excess of dehydroquinate (10000:1 substrate:enzyme) it was possible to observe a second series of signals of lower intensity (Figure 2). These correspond to a species of molecular weight  $27618 \pm 7.4$ , which is that expected for 4, the imine adduct between the enzyme and the product (calculated molecular weight 27621.9).

The low intensity of the imine adduct observed in Figure 2 is probably a consequence of the extreme conditions the liganded enzyme encounters upon injection into the spectrometer.<sup>13,15</sup> The observation of the imine adduct with the product rather than the substrate is consistent with the result from the borohydride trapping experiment and suggests that 4 is the major covalently bound intermediate on the enzyme. This is reasonable as the equilibrium constant for the dehydration is 15.<sup>16</sup>

In summary, the use of electrospray mass spectrometry has allowed direct observation of an imine intermediate in an enzyme reaction. The molecular weight of both the imine intermediate and the adduct formed after reduction with sodium borohydride suggests that the major covalent enzyme adduct is the imine with the product 3-dehydroshikimate. The inactivation experiment also showed the enzyme does not exhibit half-sites reactivity. These experiments provide a powerful demonstration of the use of electrospray mass spectrometry to study enzyme mechanisms.

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## **Externally Bound Metal Ion Complexes of** Buckminsterfullerene, $MC_{60}^+$ , in the Gas Phase

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Shortly after their discovery of the special stability of the fullerenes,<sup>1</sup> Smalley and co-workers demonstrated that metalfullerene species could be generated in the gas phase by growing them in a supersonic expansion source following laser desorption from a graphite target impregnated with various metals, M = La, K, and Cs.<sup>2,3</sup> Cox and co-workers made similar observations, but did not consider these to be fullerene derivatives.<sup>4,5</sup> As with the fullerenes themselves, the metallated species were detected mass spectrometrically and their structures probed by multiphoton photodissociation. These complexes, and in particular  $MC_{60}$  and its singly charged counterpart,  $MC_{60}^+$ , were found to be highly stable, requiring significant laser fluence to cause fragmentation by sequential  $C_2 loss.^3$  Further loss of  $C_2$  ceased at some critical even number of carbon atoms depending on the metal, such as C44La<sup>+</sup> (possibly C42La<sup>+</sup>), C44K<sup>+</sup>, and C48Cs<sup>+</sup>. These results led Smalley and co-workers to predict metal-included structures in which the central metal is enclosed in an inert carbon cage. If such species could be synthesized in macroscopic amounts, as demonstrated for  $C_{60}$  and  $C_{70}$ ,<sup>6</sup> they could exhibit important chemical and physical properties. Most notably to date is the observation that preformed  $C_{60}$  doped with potassium exhibits a superconducting transition,<sup>7</sup> although an externally bound metal-fullerene association has been demonstrated.<sup>8</sup> In this work, we extend our initial study<sup>9</sup> on  $FeC_{60}^+$  to several other metal ions and demonstrate unequivocally that metals added to preformed  $C_{60}$  generate externally bound complexes. In addition, externally bound  $LaC_{60}^{+}$  is observed to exhibit dramatically different properties than its metal-included isomer.

 $MC_{60}^+$  (M = Fe, Co, Ni, Cu, Rh, La, and VO) species were formed in a Nicolet FTMS-2000 Fourier transform mass spectrometer via a multistep sequence<sup>10</sup> initiated by laser desorption to generate M<sup>+</sup> from the pure metal targets.<sup>11</sup> The next step depended on the metal ion, but the majority of the ions including Co<sup>+</sup>, Ni<sup>+</sup>, Cu<sup>+</sup>, Rh<sup>+</sup>, and La<sup>+</sup> were then permitted to react directly with a background pressure of  $C_{60}$  heated off a solids probe at 350 °C. Using a system temperature of 250 °C, charge transfer to background potassium was found to be a problem, although K<sup>+</sup> was not observed to form  $KC_{60}^+$  under these conditions. Reducing the temperature to 150 °C, however, removed the potassium background and permitted longer trapping periods to make up for the reduced  $C_{60}$  pressure. Under these conditions, the metal ions formed MC<sub>60</sub><sup>+</sup> by direct attachment together with varying amounts of  $C_{60}^+$  by charge transfer. V<sup>+</sup> and La<sup>+</sup> form very strong bonds with oxygen<sup>12</sup> and rapidly reacted with the background gas  $(\sim 2 \times 10^{-7}$  Torr using the solids probe) to form VO<sup>+</sup> and LaO<sup>+</sup>. Only VO<sup>+</sup> was observed to directly attach to  $C_{60}$  to form the fullerene complex, but conditions probably exist favorable to the formation of  $LaOC_{60}^+$ . In order to generate  $LaC_{60}^+$ , it was necessary to suppress LaO<sup>+</sup> formation. This was accomplished by encasing the external elements of the solids probe with a flexible plastic sheath through which Ar flowed. Finally, Fe<sup>+</sup> reacts with C<sub>60</sub> predominantly by charge transfer and, thus, a background of *n*-pentane at about  $4 \times 10^{-8}$  Torr was added which yields  $Fe(C_nH_{2n})^+$  (n = 2-5) species.<sup>13</sup> These ions undergo ligand exchange reactions with  $C_{60}$  to form  $FeC_{60}^+$ . A similar procedure could be used on selected metal ions to enhance  $MC_{60}^{+}$  intensities over that observed by direct attachment.

After a total reaction period, which varied from about 0.3 to 10 s depending upon the system and the conditions, the  $MC_{60}$ was isolated by double resonance ejection pulses14 and subjected to low-energy collision-induced dissociation (CID)<sup>15</sup> using Ar as the target gas at about  $2 \times 10^{-6}$  Torr. The collision energies ranged from 30 to 130 eV, laboratory frame. CID on  $FeC_{60}^+$ ,  $CoC_{60}^+$ ,  $NiC_{60}^+$ , and  $CuC_{60}^+$  yields  $C_{60}^+$ , while  $LaC_{60}^+$  and  $VOC_{60}^+$  yield  $La^+$  and  $VO^+$ , respectively, and  $RhC_{60}^+$  yields a mixture of  $C_{60}^+$  and  $Rh^+$ , with the Rh<sup>+</sup> predominating. Figure 1 for  $LaC_{60}^+$  illustrates the type of data obtained.

Given the ionization potentials<sup>16</sup> of IP(Fe) = 7.87 eV, IP(Co)= 7.86 eV, IP(Cu) = 7.726 eV, IP(Ni) = 7.635 eV, IP(Rh) =

(12) Armentrout, P. B.; Halle, L. F.; Beauchamp, J. L. J. Am. Chem. Soc. 1981, 103, 6501.

(13) Jacobson, D. B.; Freiser, B. S. J. Am. Chem. Soc. 1983, 105, 5197.
 (14) Comisarow, M.; Parisod, G.; Grassi, V. Chem. Phys. Lett. 1978, 57,

(16) Lias, S. G.; Bartmess, J. E.; Liebman, J. F.; Holmes, J. L.; Levin, R. D.; Mallard, W. G. J. Phys. Chem. Ref. Data 1988, 17, Suppl. 1.

<sup>(15)</sup> The acidity of the carrier stream in the mass spectrometer (pH 3)<sup>13</sup> will protonate the substrate (increasing  $K_m$ ) and the active site lysine on the enzyme inhibiting imine formation. Furthermore acetate is a competitive inhibitor of dehydroquinase<sup>12</sup>

<sup>(16)</sup> S. Mitsuhashi, S.; Davis, B. D. Biochim. Biophys. Acta 1954, 15, 54-61

<sup>(1)</sup> Kroto, H. W.; Heath, J. R.; O'Brien, S. C.; Curl, R. F.; Smalley, R. E. Nature 1985, 318, 162.

<sup>(2)</sup> Heath, J. R.; O'Brien, S. C.; Zhang, Q.; Liu, Y.; Curl, R. F.; Kroto,
H. W.; Tittel, F. K.; Smalley, R. E. J. Am. Chem. Soc. 1985, 107, 7779.
(3) Weiss, F. D.; Elkind, J. L.; O'Brien, S. C.; Curl, R. F.; Smalley, R. E.
J. Am. Chem. Soc. 1988, 110, 4464.

<sup>(4)</sup> Cox, D. M.; Trevor, D. J.; Reichmann, K. C.; Kaldor, A. J. Am. Chem.

Soc. 1986, 108, 2457.

<sup>(5)</sup> Cox, D. M.; Reichmann, K. C.; Kaldor, A. J. Chem. Phys. 1988, 88, 1588.

<sup>(6)</sup> Kratschmer, W.; Lamb, L. D.; Fostiropoulos, K.; Huffman, D. R. Nature 1990, 347, 354.

<sup>(7)</sup> Hebard, A. F.; Rosseinsky, M. J.; Haddon, R. C.; Murphy, D. W.; Glarum, S. H.; Palstra, T. T. M.; Ramirez, A. P.; Kortan, A. R. Nature 1991, 350, 600.

<sup>(8)</sup> Stephens, P. W.; Mihaly, L.; Lee, P. L.; Whetten, R. L.; Huang, S.-M.;
(8) Stephens, P. W.; Mihaly, L.; Lee, P. L.; Whetten, R. L.; Huang, S.-M.;
(9) Roth, L. M.; Huang, Y.; Schwedler, J. T.; Cassady, C. J.; Ben-Amotz,
D.; Kahr, B.; Freiser, B. S. J. Am. Chem. Soc. 1991, 113, 6298.
(10) Freiser, B. S. Chemtracts 1989, 1, 65.
(11) Cody, R. B.; Burnier, R. C.; Reents, W. D., Jr.; Carlin, T. J.;
McCrery, D. A.; Lengel, R. K.; Freiser, B. S. Int. J. Mass. Spectrom. Ion Phys. 1980, 33, 37

<sup>(15)</sup> Cody, R. B.; Burnier, R. C.; Cody, R. B.; Freiser, B. S. Anal. Chem. 1982, 54, 96.



Figure 1. Plot of relative abundance vs m/z: (A) isolated LaC<sub>60</sub><sup>+</sup>; (B) collision-induced dissociation of  $LaC_{60}^+$  (105 eV Laboratory, 4.7 eV center of mass energy) yielding La<sup>+</sup>.

7.46 eV, IP(VO) = 7.23 eV (Weisshaar, J. C., private communication) and IP(La) = 5.577 eV, the CID results indicate IP(Fe, Co, Ni, Cu) >  $IP(C_{60}) \sim IP(Rh) > IP(VO, La)$ , which is in agreement with a recently reported value of  $IP(C_{60}) = 7.61 \pm$ 0.11 eV.<sup>17</sup> Furthermore, these results provide strong support for internally bound "egg shell"  $MC_{60}^+$  species (particularly for M = La) reported earlier grown in a supersonic expansion, since these latter species are highly stable and lose C2 molecules when sufficiently activated. In direct contrast, the externally bound isomers, presumably always formed by adding the metal to the preformed  $C_{60}$ , require relatively little activation energy to cleave the metal from the complex, leaving an intact  $C_{60}$  species. Although the type of activation used in the current study (collisional) and that reported in the previous study (photoexcitation) can yield different fragmentation processes,<sup>18</sup> it is likely that some metal loss would be observed for an externally bound complex regardless of the method of excitation employed.

Studies are underway to synthesize other members of this new family of ions and to characterize their subsequent chemistry with other reagent gases. It should be possible to synthesize virtually any metal ion-C<sub>60</sub> complex, as well as an endless variety of ligated metal ion-C<sub>60</sub> complexes, in the gas phase. Determination of the  $M^+-C_{60}$  bond energies will also be possible by monitoring lig9419

and-exchange reactions using reference ligands.

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## Surface Electrostatics, Reduction Potentials, and the **Internal Dielectric Constant of Proteins**

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Electrostatic contributions by surface charges are presumed to play important roles in dictating the affinity and selectivity of numerous protein-macromolecule and protein-small molecule interactions. Although numerous modeling and theoretical efforts have attempted to predict the detailed electrostatic fields in and around globular proteins,<sup>1</sup> there have been only a few systematic efforts to experimentally map the electrostatic potential surface of a protein in solution and correlate this with simplified theories.<sup>1h</sup> In this communication we demonstrate that site-directed mutagenesis of surface charges together with high-resolution electrochemical measurements can be used to quantitate the boundary-value electrostatic potentials of soluble proteins.

Prosthetic group reduction potentials<sup>2</sup> and amino acid side chain ionization constants can be affected by surface-charged residues.<sup>3</sup> Here we show that precise measurement of the shift in the reduction potential for a heme prosthetic group upon alteration of surface charge can be used to test the validity of theoretical calculations of macromolecular electrostatic fields and, in a simple two-continuum dielectric model, to estimate the effective bulk internal dielectric constant of a protein. By using amino acid replacements at numerous sites on the surface, we find that a single distance-dependent dielectric will not fit experimental data, but rather the detailed shape of the macromolecule must be taken into account.

We have chosen rat liver cytochrome  $b_5$  for these investigations, since previous studies have shown that the protein is structurally rigid to surface mutations.<sup>4</sup> The heme redox potential was measured by direct electrochemical methods using a cysteinemodified gold working electrode.<sup>5</sup> For these investigations we

(4) (a) Rodgers, K. K.; Pochapsky, T. C.; Sligar, S. G. Science 1988, 240,

(5) (a) Di Gleria, K.; N.; N.; K.; Sligar, S. G. J. Mol. Biol., in press.
(5) (a) Di Gleria, K.; Hill, H. A. O.; Lowe, V. J.; Page, D. J. J. Electroanal. Chem. 1986, 213, 333–338. (b) Hill, H. A. O.; Lawrance, G. A. J. Electroanal. Chem. 1989, 270, 309–318. (c) Bagby, S.; Barker, P. D.; Guo, L.-H.; Hill, H. A. O. Biochemistry 1990, 29, 3213–3219.

<sup>(17)</sup> Zimmerman, J. A.; Eyler, J. R.; Bach, S. B. H.; McElvaney, S. W. J. Chem. Phys. 1991, 95, 3267.
(18) Franchetti, V.; Freiser, B. S.; Cooks, R. G. Org. Mass Spectrom.

<sup>1978, 13, 106.</sup> 

<sup>\*</sup>To whom correspondence should be addressed.

<sup>(1) (</sup>a) Mathew, J. B. Annu. Rev. Biophys. Biophys. Chem. 1985, 14, 387-417. (b) Honig, B. H.; Hubbell, W. L.; Flewelling, R. F. Annu. Rev. Biophys. Biophys. Chem. 1986, 15, 163-193. (c) Schoefer, M.; Froemmel, C. J. Mol. Biol. 1990, 216, 1045-1066. (d) Bashford, D. Curr. Opin. Struct. Biol. 1991, 1, 175-184. (e) Harvy, S. C. Proteins 1989, 5, 78-92. (f) Sharp, K. A.; Honig, B. Annu. Rev. Biophys. Biophys. Chem. 1990, 19, 301-332. (g) Davis, M. E.; McCammon, J. A. Chem. Rev. 1990, 90, 509-521. (h) Bashford, D.; Karplus, M. *Biochemistry* **1990**, *29*, 10219-10225. (i) Karshikov, A. D.; Engh, R.; Bode, W.; Atanasov, B. P. Eur. Biophys. J. **1989**, *17*, 287-297.

<sup>(2) (</sup>a) Smith, H. T.; Staudenmeyer, M.; Millett, F. Biochemistry 1977, 16, 4971-4974. (b) Reid, L. S.; Mauk, M. R.; Mauk, A. G. J. Am. Chem. Soc. 1984, 106, 2182-2185. (c) Caffrey, M. S.; Cusanovich, M. A. Arch. Biochem. Biophys. 1991, 285, 227-230.

<sup>(3)</sup> Sternberg, M. J. E.; Hays, F. R. F.; Russell, A. J.; Thomas, P. G.; Fersht, A. R. Nature 1987, 330, 86–88.